

# Analysis and Comparison of Human Prostasomes Amino Acid Content Variation in Normal Men and Infertile Men-A Clinical Relevance Study for Effective Diagnosis Method for Human Male Infertility

D. Vignesh<sup>1</sup>; Dr.A.S. Vickram<sup>2\*</sup>

<sup>1</sup>Research Scholar, Department of Biomedical Engineering, Saveetha School of Engineering, Saveetha Institute of Medical and Technical Sciences, Saveetha University, Chennai, Tamil Nadu, India. <sup>1</sup>vigneshdanushkodi17@saveetha.com

<sup>2\*</sup>Project Guide, Department of Biotechnology, Saveetha School of Engineering, Saveetha Institute of Medical and Technical Sciences, Saveetha University, Chennai, Tamil Nadu, India.

<sup>2\*</sup>vickramas.sse@saveetha.com

# Abstract

Aim: The main objective of this study is to compare and analyse human prostasomes amino acid content variation in normal men semen samples and infertile men semen samples for identification of clinical relevance. **Materials and methods:** Semen samples were collected from normal men (N=32) and from infertile men (N=32) and by following the standard world health organisation protocol semen analysis was done. Amino acid quantification was done by using amino acid analyzer. Prostasomes were separated from spermatozoa and seminal plasma by using centrifugation technique at 95000 RPM for 90 mins. **Results:** Independent sample T-test was carried out and shows that proline and alanine amino acids concentration (p<0.01) statistically significant compared with fertile men and infertile men. High concentration of amino acids in prostasomes were found in fertile men samples  $(18.09 \pm 0.20 \ \mu moles/L)$  when compared with infertile men samples  $(15.12\pm 0.37 \ \mu moles/L)$ . **Conclusion:** Amino acid in prostasomes plays an important role in the fertilization; the change in the concentration of amino acid in prostasomes leads to infertility of men. Here we found that the concentration of amino acids is high in fertile men when compared to infertile men which could act as an innovative diagnosis method for infertility.

**Key-words:** Amino Acids, Prostasomes, Amino Acid Analyzer, Infertility, Innovative Methods for Diagnosis, Semen, Centrifugation, Reproductive Medicine.

# 1. Introduction

This research is about to identify the clinical relevance of amino acid content variation in human prostasomes of fertile and infertile men. Male Infertility rate has been increased about 30% in the past 10 years. Male infertility is due to various reasons that occurs in the semen and its parameters. The importance of this research is to keep evaluating the amino acid contents in prostasomes for diagnosis of human male infertility. Worldwide these types of research become important as diagnosis of male infertility needs more attention (A.S. Vickram, Samad, et al. 2020). Seminal fluid components that present in the semen varies with people due to several conditions like temperature, climate and food habit (A.S. Vickram, Samad, et al. 2020). Prostasomes are the extracellular fluid which have gained a huge attention due to its sufficient cause such as clear access in the semen fluids (György et al. 2011). These prostasomes are termed as the extracellular membrane fluids which usually ranges from 40-5000mm in diameter. They are categorized based on their origin, size, morphology, and its mode of ejection (Witwer et al. 2013). This study results may lead to application in the area of andrology and reproductive medicine.

We looked for the most cited articles in the pubmed and science direct database whereas it ended with 425 articles published in this domain. Amino acid concentration in prostasomes and seminal plasma is one of the important parameters which plays a key role in deciding fertility status of men. The most cited article describes that variation in seminal fluid profile can be representative of genital tract dysfunctions and thus serve as an infertility biomarker (Herwig et al. 2013). In this regard, while exosomes isolated from seminal plasma of asthenospermia and azoospermic patients have similar form, scale, and expression of typical normospermic patients differs from the infertile men whereas 50% of infertility cases is due to content variation in seminal fluid (Candenas and Chianese 2020). Approximately 2/3 of infertile men have a sperm production problem, which includes a low sperm count, poor sperm parameters, and a high non-motile sperm count (Carrell et al. 2016). Infertility is on the rise with 25% of couples attempting but failing to conceive. The male factor is responsible for more than 40% (Sengupta 2015). To make the testing process simple here a novel technique is approached that amino acid concentration variation in prostasomes of normal men and infertile men.

Previously our team has a rich experience in working on various research projects across multiple disciplines (Sathish and Karthick 2020; Varghese, Ramesh, and Veeraiyan 2019; S. R. Samuel, Acharya, and Rao 2020; Venu, Raju, and Subramani 2019; M. S. Samuel et al. 2019; Venu,

Subramani, and Raju 2019; Mehta et al. 2019; Sharma et al. 2019; Malli Sureshbabu et al. 2019; Krishnaswamy et al. 2020; Muthukrishnan et al. 2020; Gheena and Ezhilarasan 2019; Vignesh et al. 2019; Ke et al. 2019; Vijayakumar Jain et al. 2019; Jose, Ajitha, and Subbaiyan 2020). Now the growing trend in this area motivated us to pursue this project.

There is no clinical relevance study for variation of amino acids content in prostasomes of fertile men and infertile men. We had already expertised in this field of research for over a decade. The major aim of this current study is to analyse and compare the prostasomes concentration in the seminal fluid of fertile men and infertile men. The concentration of amino acids in prostasomes is high in fertile men when compared to infertile men.

# 2. Materials and Methods

This study was conducted at biochemistry lab in saveetha school of engineering. Samples were collected in accordance with the world organization (WHO) standard procedure. Sample size was collected by using previous study results (García-Rodríguez et al. 2018) in clinicalc.com by keeping threshold 0.05 and G power 80%, confidence interval 95% and enrollment ratio as 1. Two different groups were taken for the analysis: one is a fertile men group (N=32) and the other one is infertile men group (N=32). Computer assisted semen analysis (CASA)- german made and amino acid analyser were used in this study for analysis.

The semen samples which are used for this research were obtained from the milan fertility center, bangalore, karnataka. The samples are collected from the people who are in abstinence time (about 4 to 7 days) and then recorded. The samples are collected through a mastrubation process in a clean and intoxic wide mounted plastic container in the sample collection room. The liquefaction of samples are done and time is noted (A.S. Vickram, Anbarasu, et al. 2020) Computer assisted semen analysis is a modern technique which differs from the manual semen analysis by the process of evaluation (Agarwal, Henkel, and Majzoub 2021) The modern CASA systems are designed in a way of measuring quantitatively the several aspects of prostasomes content such as sperm concentration, sperm motility, and its morphology through this CASA technique the semen parameters are identified for both the fertile group and infertile group.

The step-in separation of prostasomes from amino acid is centrifugation. Centrifugation operates on the idea that two particles in suspension (cells, organelles, or molecules) of different masses or densities can settle at different rates to the bottom of a tube (Li and Boix 2021). 800 RPM

separates the sperm cells in 8 minutes under 4°c, 1000 RPM separates the debris in 10 minutes under 4°c and 95,000 RPM for prostasomes in 90 minutes under 4°c.

### **Statistical Analysis**

The statistical comparison of fertile men group and infertile men group was done through SPSS version 21. There are no dependent variables whereas the independent variables are prostasomes and amino acids volume, sperm motility. Analysis was done for mean, standard deviation, independent T-test.

# 3. Results

Semen analysis was performed and reported in Table 1 for major parameters like volume, pH, sperm concentration, total motility, rapid progressive motility, and normal morphology, all the parameters were shown with normal values as per world health organization in case of normospermia and the values where not compatible with the world health organization in case of all infertile conditions which reflected in Table 1.

Table 1- Represents the Mean ± Standard Error for major Semen Parameters between Fertile Men and Infertile Men. From the<br/>Table it can be Observed that the Infertile Conditions such as Oligoasthenospermia, Oligospermia, Azoospermia and<br/>Asthenospermia have very Low Volume (ml), low pH, Low Sperm Concentration (millions/ml) when compared to the<br/>Normospermia. The Normal Morphology of Infertile Group is very Low with 12.54 % in Oligoasthenospermia whereas in the<br/>Fertile Group it was High with 40.3 %

Semen category	Volume (ml)	рН	Sperm concentration (millions/ml)	Total motility (%)	Rapid progressive Motility (%)	Normal morphology (%)	
Oligoasthenospermia (N=8)	2.9±0.8	7.7±06.2	4.6±0.4	5.3±1.4	2.02±0.6	17.2±2.3	
Asthenospermia (N=8)	2.3±0.2	7.7±0.2	28.8±4.2	08.3±1.8	4.6±1.6	12.54±1.4	
Azoospermia (N=8)	2.2±0.3	7.8±0.2	NIL	NIL	NIL	NIL	
Normospermia (N=16)	3.3±0.7	7.8±0.3	85.7±7.3	42.31±10.2	28.4±5.4	22.3±3.9	
Oligospermia (N=8)	2.6±0.5	7.6±0.2	7.3±0.9	19.9±2.4	20.5±5.7	22.6±3.2	
Control (N=16)	3.6±0.9	7.8±0.1	89.4±15.1	48.7±6.6	31.1±4.9	40.3±4.4	

Prostasomes content in mg/ml was evaluated for all the infertile conditions and fertile conditions and depicted in Table 2. In Table 2, it was observed that the mean value for prostasomes content was found to be 2.46 in control which was 3 times higher than oligoasthenospermia.

Table 2- Represent the Prostasomes Content (mg/ml) for various Infertile Conditions and Fertile Conditions. From this Table it was observed that Infertile Groups have Low Prostasomes Concentration when Compared with Fertile Men Semen Samples. The Control Group has a High Mean Value with 2.46±0.19 (mg/ml) whereas the Infertile Category Azoospermia have Low mean with 0.09±0.22 (mg/ml)

category	Prostasomes (mg/ml)
Oligospermia (N=8)	0.92±0.09
Oligoasthenospermia (N=8)	0.83±0.21
asthenospermia(N=8)	0.51±0.16
azoospermia(N=8)	0.09±0.22
normospermia(N=16)	2.12±0.25
control(N=16)	2.46±0.19

From Table 3, it was observed that the fertile group has high concentration of amino acid except serine when compared to the infertile group. 13 amino acids concentration are identified through the amino acid analyser which have shown a drastic difference in the fertile group and in infertile group that are depicted in Table 3. The mean amino acid content in the fertile group is 18.09 ( $\mu$  moles/ L) and in infertile category it is about 15.12 ( $\mu$  moles/ L), this shows the difference in between fertile and infertile categories shown in Table 4. Comparison of amino acid content in prostasomes was done in Table 5 between fertile and infertile men, the independent T test was done and found that amino acids proline and alanine was found with significant difference between fertile and infertile category.

Table 3- Represents the Amino Acids that are Quantified in the Prostasomes in Fertile Semen Samples and in Infertile Semen Samples. From this Table it was Observed that the Fertile Group has High Concentration of Amino Acid except Serine when Compared to the Infertile Group. 13 Amino Acids Concentration are Identified through the Amino Acid Analyser which have shown a Drastic difference in the Fertile Group and in Infertile Group. **NS** – difference in Mean is Insignificant,\*\* Signifies p<0.01,\*\*\* signifies p<0.001

Infertile Group (µ moles/ L)	Control Group (Fertile) (µ moles/ L)	Amino acid
$0.61 \pm 0.04$	4.52 ± 0.18 ***	Aspartic acid
$2.17 \pm 0.01$	5.73 ± 0.18 **	Tyrosine
$1.75 \pm 0.29$	$1.53\pm0.18^{-NS}$	Valine
$2.84 \pm 0.13$	$2.52 \pm 0.10^{-NS}$	Asparagine
$1.23 \pm 0.07$	$1.54 \pm 0.11$ <sup>NS</sup>	Glutamine
$18.03 \pm 0.24$	4.56 ± 0.38 ***	Serine
$1.86 \pm 0.15$	9.41 ± 0.13 ***	Glycine
$3.91\pm0.07$	$1.14 \pm 0.05 **$	Phenylalanine
$0.92 \pm 0.07$	$0.93 \pm 0.0.4$ <sup>NS</sup>	Amino butyric acid
$1.11 \pm 0.13$	3.29 ± 0.06 ***	Glutamic Acid
$5.05 \pm 0.14$	2.42 ± 0.12 ***	Alanine
$0.92 \pm 0.06$	2.05 ± 0.02 ***	Histidine
$0.721 \pm 0.03$	2.04 ± 0.07 **	Proline

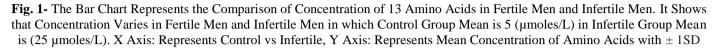
Table 4- Represents the Total Amino Acid Content in Fertile and Infertile Prostasomes Fraction. In which each Category Contains 32 Samples. The Mean Amino Acid Content in Fertile Group is 18.09 (μ moles/ L) and in Infertile Category it is about 15.12 (μ Moles/ L)

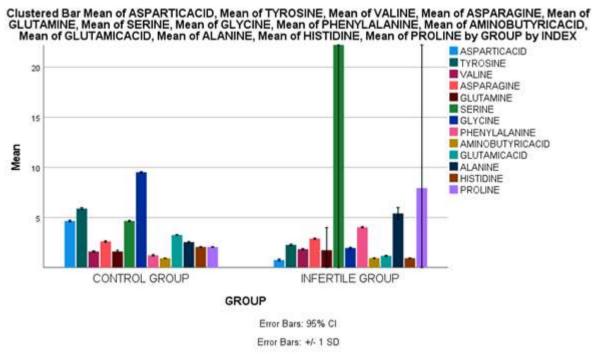
	Fertile category	Infertile category
No of samples	32	32
Amino acid content (µ moles/ L)	18.0±90.20	15.12±0.37

<b>Table 5-</b> Represents that Independent Sample T-Test which Shows the Significance in which the Amino Acids such as Alanine,
Proline are found with Statistical Significance (p<0.01) when Comparing Fertile Groups with Infertile Groups

		Leven's test for equality of variances				t-test for equality of means		95% confidence Interval of the difference		
		F	Sig	t	df	Sig [2- tailed]	Mean diff.	Std. Error diff.	Lower	Upper
Aspartic Acid	Equal variances assumed	.497	.483	181.18	62	<.001	3.89	.021	3.85	3.93
	Equal variances not assumed			181.18	61.37	<.001	3.89	.021	3.85	3.93
Tyrosine	Equal variances assumed	4.48	0.38	172.49	62	<.001	3.62	.021	3.58	3.66
	Equal variances not assumed			172.49	56.55	<.001	3.62	.021	3.58	3.66
Valine	Equal variances assumed	.020	.889	- 14.845	62	<.001	-232	.016	263	2
	Equal variances not assumed			- 14.845	61.91	<.001	23	.016	263	20
Asparagine	Equal variances assumed	1.27	.262	- 10.239	62	<.001	28	.015	310	24
	Equal variances not assumed			- 10.239	59.59	<.001	28	.015	310	24
Glutamine	Equal variances assumed	3.46	.067	290	62	.773	11	.394	901	.673
Giutannie	Equal variances not assumed			290	31.18	.774	11	.394	917	.689
Serine	Equal variances assumed	4.26	.043	-1.241	62	.219	-69.499	55.995	-181.431	42.434
	Equal variances not assumed			-1.241	31.00	.224	-69.499	55.995	-183.701	44.704
Glycine	Equal variances assumed	1.34	.25	452.41	62	<.001	7.56	.017	7.5	7.59
	Equal variances not assumed			452.41	60.58	<.001	7.56	.017	7.52	7.59
Phenylalanine	Equal variances assumed	1.92	.170	- 168.89	62	<.001	-2.8	.017	-2.83	-2.7
Thenykululline	Equal variances not assumed			- 168.89	60.76	<.001	-2.8	.017	-2.83	-2.7
Aminobutyric	Equal variances assumed	11.2	.001	-1.354	61	.178	00	.006	019	.004
Acid	Equal variances not assumed			-1.373	53.01	.176	00	.006	019	.004
Glutamic Acid	Equal variances assumed	5.05	.028	200.25	62	<.001	2.08	.010	2.06	2.10
	Equal variances not assumed			208.25	55.48	<.001	2.08	.010	2.06	2.10
Alanine	Equal variances assumed	16.2	<.001	- 27.595	61	<.001	-2.8	.104	-3.08	-2.6
	Equal variances not assumed			- 27.167	30.93	<.001	-2.8	.106	-3.09	-2.6
Histidine	Equal variances assumed	.001	.980	169.29	62	<.001	1.11	.007	1.10	1.13
	Equal variances not assumed			169.29	61.99	<.001	1.11	.007	1.10	1.13
Proline	Equal variances assumed	15.8	<.001	-1.454	62	.151	-5.6	3.887	-13.4	2.12
1 round	Equal variances not assumed			-1.454	31.00	.156	-5.6	3.887	-13.5	2.27

13 different amino acids content were compared in Fig. 1 and found that proline and alanine yielded better results in case of fertile men, It shows that concentration varies in fertile men and infertile men in which control group mean is 5 ( $\mu$ moles/L) in infertile group mean is (25  $\mu$ moles/L).





### 4. Discussion

Our overall results show that there are huge variations observed in the concentration of amino acids present in human prostatosmes between various infertile conditions and normospermia men. However the identification of the sperm and amino acid concentration as an ideal biomarker is a major challenge, our results were found to be in accordance with the studies conducted by Agrawal et al (Agarwal, Selvam, and Baskaran 2020). Due to the lack of alternative testing or the need for invasive diagnostic procedures, many clinical areas in the field of male fertility are primed for the production of seminal biomarkers, our results may lead to use of prostasomes as one of the biomarker for male infertility, and all our results in accordance with Bieniek (Bieniek, Drabovich, and Lo 2016). Prostasomes present less in infertile category in our study, more similar findings were observed in the study conducted by Garcia et al (García-Rodríguez et al. 2018).

We followed the standard protocol of the World Health Organization, 2010 very strictly, for the preparation of semen analysis report, we segregated the infertile groups only based on the standard values mentioned, in our study, got better results for semen analysis report. Amino acid analyzer and CASA instruments were calibrated completely before analysis for better results. Semen samples were collected in a toxic-free plastic container, so that the sperm could survive even after collection for correct analysis. Samples were analysed by already standardized protocol by vickram et al (S. Vickram et al. 2021) (A.S. Vickram, Anbarasu, et al. 2020) (S. Vickram et al. 2021). The prostasomes which are present in the semen is an excellent source of biomarker which makes it an innovative method of diagnosis for male infertility (Ebert, Kisiela, and Maser 2015). The analysis of amino acid level in the fertile group and infertile group from that we could identify that amino acids in the fertile group is high in concentration whereas in infertile group the concentration of amino acid is low. In 13 amino acids all amino acids have drastic variation when comparing them with other groups.

Our institution is passionate about high quality evidence based research and has excelled in various fields ((Vijayashree Priyadharsini 2019; Ezhilarasan, Apoorva, and Ashok Vardhan 2019; Ramesh et al. 2018; Mathew et al. 2020; Sridharan et al. 2019; Pc, Marimuthu, and Devadoss 2018; Ramadurai et al. 2019). We hope this study adds to this rich legacy.

We had some limitations in our study execution, while using amino acid analyzer turbidity is an issue due to the particulates in the samples, CASA had limitations in guaranteeing the identification of sperm parameters such as motility and concentration for continuous examination. Centrifugation for 1 hour to separate the prostomses from seminal plasma was another limitation as the machine got heat and not maintaining -4 degree celsius for continuous time, this may lead to denaturation of protein.

Still the relevance and few more properties of prostasomes and amino acids remains unknown. Vast development and multi omics approach on prostasomes will lead the researchers to focus on new findings on seminal fluid with unique properties that can be identified in a better way.

# 5. Conclusion

The concentration of amino acid present in prostasomes of fertile men is high about 18.09  $\pm$  0.20 (µ moles/ L) when compared to infertile men 15.12 $\pm$  0.37 (µ moles/ L). This identification led to the conclusion that drastic reduction in amino acids concentration of prostasomes will lead to

infertility. Prostasomes play an important role for infertility and this could be used for the diagnosis purpose.

### Declarations

### **Conflict of interests**

No conflict of interests in this manuscript.

### **Authors Contributions**

Author DV was involved in data collection, data analysis, manuscript writing. Author VAS was involved in conceptualization, data validation, and critical review of manuscript.

### Acknowledgments

The authors would like to express their gratitude towards Saveetha School of engineering, Saveetha Institute of Medical and Technical Sciences (Formerly known as Saveetha University) for providing the necessary infrastructure to carry out this work successfully.

### Funding

We thank the following organizations for providing financial support that enabled us to complete the study.

- 1. BCX Bio Organics, Bangalore, India.
- 2. Saveetha University
- 3. Saveetha Institute of Medical and Technical Sciences.
- 4. Saveetha School of engineering.

### References

Agarwal, A., Henkel, R., & Majzoub, A. (Eds.). (2021). *Manual of Sperm Function Testing in Human Assisted Reproduction*. Cambridge University Press.

Agarwal, A., Panner Selvam, M.K., & Baskaran, S. (2020). Proteomic analyses of human sperm cells: understanding the role of proteins and molecular pathways affecting male reproductive health. *International journal of molecular sciences*, *21*(5), 1621. https://doi.org/10.3390/ijms21051621

Bieniek, J.M., Drabovich, A.P., & Lo, K.C. (2016). Seminal biomarkers for the evaluation of male infertility. *Asian journal of andrology*, *18*(3), 426. https://doi.org/10.4103/1008-682x.175781

Candenas, L., & Chianese, R. (2020). Exosome composition and Seminal Plasma Proteome: A promising source of biomarkers of male infertility. *International Journal of Molecular Sciences*, 21(19), 7022. https://doi.org/10.3390/ijms21197022

Carrell, D.T., Aston, K.I., Oliva, R., Emery, B.R., & De Jonge, C.J. (2016). The "omics" of human male infertility: integrating big data in a systems biology approach. Cell and tissue research, 363(1), 295-312. https://doi.org/10.1007/s00441-015-2320-7

Ebert, B., Kisiela, M., & Maser, E. (2015). Human DCXR–another 'moonlighting protein'involved in sugar metabolism, carbonyl detoxification, cell adhesion and male fertility? *Biological Reviews*, 90(1), 254-278.

Ezhilarasan, D., Apoorva, V.S., & Ashok Vardhan, N. (2019). Syzygium Cumini Extract Induced Reactive Oxygen Species-Mediated Apoptosis in Human Oral Squamous Carcinoma Cells" *Journal of Oral Pathology & Medicine: Official Publication of the International Association of Oral Pathologists and the American Academy of Oral Pathology*, 48(2), 115-121.

García- Rodríguez, A., De la Casa, M., Peinado, H., Gosálvez, J., & Roy, R. (2018). Human prostasomes from normozoospermic and non-normozoospermic men show a differential protein expression pattern. *Andrology*, *6*(4), 585-596.

Gheena, S., & Ezhilarasan, D. (2019). Syringic acid triggers reactive oxygen species-mediated cytotoxicity in HepG2 cells. *Human & experimental toxicology*, *38*(6), 694-702.

György, B., Szabó, T.G., Pásztói, M., Pál, Z., Misják, P., Aradi, B., & Buzás, E.I. (2011). Membrane vesicles, current state-of-the-art: emerging role of extracellular vesicles. *Cellular and molecular life sciences*, *68*(16), 2667-2688.

Herwig, R., Knoll, C., Planyavsky, M., Pourbiabany, A., Greilberger, J., & Bennett, K.L. (2013). Proteomic analysis of seminal plasma from infertile patients with oligoasthenoteratozoospermia due to oxidative stress and comparison with fertile volunteers. *Fertility and sterility*, *100*(2), 355-366.

Jose, J., & Subbaiyan, H. (2020). Different treatment modalities followed by dental practitioners for ellis class 2 fracture–A questionnaire-based survey. *The Open Dentistry Journal*, *14*(1), 59–65.

Ke, Y., Al Aboody, M.S., Alturaiki, W., Alsagaby, S.A., Alfaiz, F.A., Veeraraghavan, V.P., & Mickymaray, S. (2019). Photosynthesized gold nanoparticles from Catharanthus roseus induces caspase-mediated apoptosis in cervical cancer cells (HeLa). *Artificial cells, nanomedicine, and biotechnology*, 47(1), 1938-1946.

Krishnaswamy, H., Muthukrishnan, S., Thanikodi, S., Antony, G.A., & Venkatraman, V. (2020). Investigation of air conditioning temperature variation by modifying the structure of passenger car using computational fluid dynamics. *Thermal Science*, *24*(1 Part B), 495-498.

Li, J., & Boix, E. (2021). Host Defence RNases as Antiviral Agents against Enveloped Single Stranded RNA Viruses. *Virulence*, 12(1), 444-469.

Malli Sureshbabu, N., Selvarasu, K., Nandakumar, M., & Selvam, D. (2019). Concentrated growth factors as an ingenious biomaterial in regeneration of bony defects after periapical surgery: A report of two cases. *Case reports in dentistry*.

Mathew, M.G., Samuel, S.R., Soni, A.J., & Roopa, K.B. (2020). Evaluation of adhesion of Streptococcus mutans, plaque accumulation on zirconia and stainless steel crowns, and surrounding gingival inflammation in primary molars: Randomized controlled trial. *Clinical oral investigations*, 24(9), 3275-3280. https://link.springer.com/article/10.1007/s00784-020-03204-9.

Mehta, M., Tewari, D., Gupta, G., Awasthi, R., Singh, H., Pandey, P., & Satija, S. (2019). Oligonucleotide therapy: an emerging focus area for drug delivery in chronic inflammatory respiratory diseases. *Chemico-biological interactions*, *308*, 206-215.

Muthukrishnan, S., Krishnaswamy, H., Thanikodi, S., Sundaresan, D., & Venkatraman, V. (2020). Support vector machine for modelling and simulation of Heat exchangers. *Thermal Science*, *24*(1 Part B), 499-503.

PC, J., Marimuthu, T., Devadoss, P., & Kumar, S.M. (2018). Prevalence and measurement of anterior loop of the mandibular canal using CBCT: A cross sectional study. *Clinical implant dentistry and related research*, 20(4), 531-534. https://europepmc.org/article/med/29624863

Ramadurai, N., Gurunathan, D., Samuel, A.V., Subramanian, E., & Rodrigues, S.J. (2019). Effectiveness of 2% Articaine as an anesthetic agent in children: randomized controlled trial. *Clinical oral investigations*, 23(9), 3543-3550.

Ramesh, A., Varghese, S., Jayakumar, N.D., & Malaiappan, S. (2018). Comparative estimation of sulfiredoxin levels between chronic periodontitis and healthy patients–A case-control study. *Journal of periodontology*, 89(10), 1241-1248.

Samuel, M.S., Bhattacharya, J., Raj, S., Santhanam, N., Singh, H., & Singh, N.P. (2019). Efficient removal of Chromium (VI) from aqueous solution using chitosan grafted graphene oxide (CS-GO) nanocomposite. *International journal of biological macromolecules*, *121*, 285-292.

Samuel, S.R., Acharya, S., & Rao, J.C. (2020). School Interventions–based Prevention of Early-Childhood Caries among 3–5-year-old children from very low socioeconomic status: Two-year randomized trial. *Journal of public health dentistry*, 80(1), 51-60.

Sathish, T., & Karthick, S. (2020). Wear behaviour analysis on aluminium alloy 7050 with reinforced SiC through taguchi approach. *Journal of Materials Research and Technology*, *9*(3), 3481-3487.

Sengupta, P. (2015). Reviewing reports of semen volume and male aging of last 33 years: From 1980 through 2013. *Asian Pacific Journal of Reproduction*, *4*(3), 242-246.

https://doi.org/10.1016/j.apjr.2015.06.010

Sharma, P., Mehta, M., Dhanjal, D.S., Kaur, S., Gupta, G., Singh, H., & Satija, S. (2019). Emerging trends in the novel drug delivery approaches for the treatment of lung cancer. *Chemico-biological interactions*, *309*, 108720.

Sridharan, G., Ramani, P., Patankar, S., & Vijayaraghavan, R. (2019). Evaluation of salivary metabolomics in oral leukoplakia and oral squamous cell carcinoma. *Journal of Oral Pathology & Medicine: Official Publication of the International Association of Oral Pathologists and the American Academy of Oral Pathology* 48 (4): 299–306.

Varghese, S.S., Ramesh, A., & Veeraiyan, D.N. (2019). Blended Module-Based Teaching in Biostatistics and Research Methodology: A Retrospective Study with Postgraduate Dental Students. *Journal of dental education*, 83(4), 445-450.

Venu, H., Raju, V.D., & Subramani, L. (2019). Combined effect of influence of nano additives, combustion chamber geometry and injection timing in a DI diesel engine fuelled with ternary (diesel-biodiesel-ethanol) blends. *Energy*, *174*, 386-406.

Venu, H., Subramani, L., & Raju, V.D. (2019). Emission reduction in a DI diesel engine using exhaust gas recirculation (EGR) of palm biodiesel blended with TiO2 nano additives. *Renewable Energy*, *140*, 245-263.

Vickram, A.S., Anbarasu, K., Gulothungan, G., Thanigaivel, S., Nanmaran, R., & Palanivelu, J. (2020). Characterization of human prostasomes protein Clusterin (macromolecule)–a novel biomarker for male infertility diagnosis and prognosis. *Journal of Biomolecular Structure and Dynamics*, 1-10. https://doi.org/10.1080/07391102.2020.1852960

Vickram, A.S., Samad, H.A., Latheef, S.K., Chakraborty, S., Dhama, K., Sridharan, T.B., & Gulothungan, G. (2020). Human prostasomes an extracellular vesicle–Biomarkers for male infertility and prostrate cancer: The journey from identification to current knowledge. *International journal of biological macromolecules*, *146*, 946-958. https://doi.org/10.1016/j.ijbiomac.2019.09.218

Vickram, S., Rohini, K., Srinivasan, S., Nancy Veenakumari, D., Archana, K., Anbarasu, K., & Srikumar, P. S. (2021). Role of Zinc (Zn) in Human Reproduction: A Journey from Initial Spermatogenesis to Childbirth. *International journal of molecular sciences*, 22(4), 2188. https://doi.org/10.3390/ijms22042188.

Vignesh, R., Ditto Sharmin, C., Annamalai, S., & Baghkomeh, P. N. (2019). Management of complicated crown-root fracture by extra-oral fragment reattachment and intentional reimplantation with 2 years review. *Contemporary clinical dentistry*, *10*(2), 397–401.

Jain, S.V., Muthusekhar, M.R., Baig, M.F., Senthilnathan, P., Loganathan, S., Wahab, P.A., & Vohra, Y. (2019). Evaluation of three-dimensional changes in pharyngeal airway following isolated lefort one osteotomy for the correction of vertical maxillary excess: a prospective study. *Journal of maxillofacial and oral surgery*, *18*(1), 139-146.

Vijayashree Priyadharsini, J. (2019). In silico validation of the non-antibiotic drugs acetaminophen and ibuprofen as antibacterial agents against red complex pathogens. *Journal of periodontology*, *90*(12), 1441-1448.

Witwer, K.W., Buzás, E.I., Bemis, L.T., Bora, A., Lässer, C., Lötvall, J., & Hochberg, F. (2013). Standardization of sample collection, isolation and analysis methods in extracellular vesicle research. *Journal of extracellular vesicles*, *2*(1), 20360. https://doi.org/10.3402/jev.v2i0.20360